

Identifying and tracking bacterial strains in metagenomic libraries

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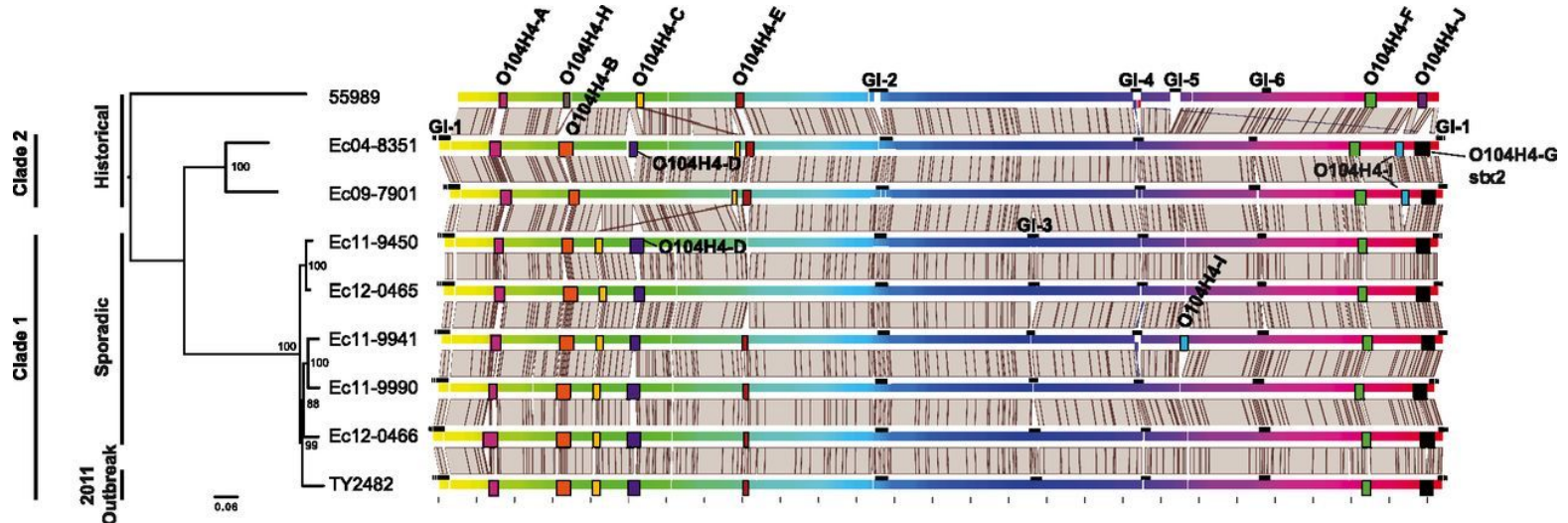
NIH T32 DK007007

The human microbiome is important and diverse

- Involved in numerous physiological processes:
 - Digestion
 - Immune regulation
 - Pathogen resistance
- Complex community
 - Highly variable across individuals
 - Hundreds of bacterial species
 - Huge but under-explored diversity *within* species

Diversity within species (strains)

- Variation in gene sequences and content within species
- Relevant effects:
e.g. Antibiotic resistance, Drug metabolism, Immune evasion, Toxin production



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e.g. Antibiotic resistance, Drug metabolism, Immune evasion, Toxin production
- Intraspecific diversity is not captured by SOP taxonomic surveys
 - 16S rRNA gene evolves too slowly

How to survey bacterial strains?



- Culturing is the “Gold Standard”
- Enables phys. characterization
- Low-throughput, expensive, biased



- Metagenomics
- Easy, high-throughput, less biased
- Interpretation often difficult

Identifying strains in metagenomes

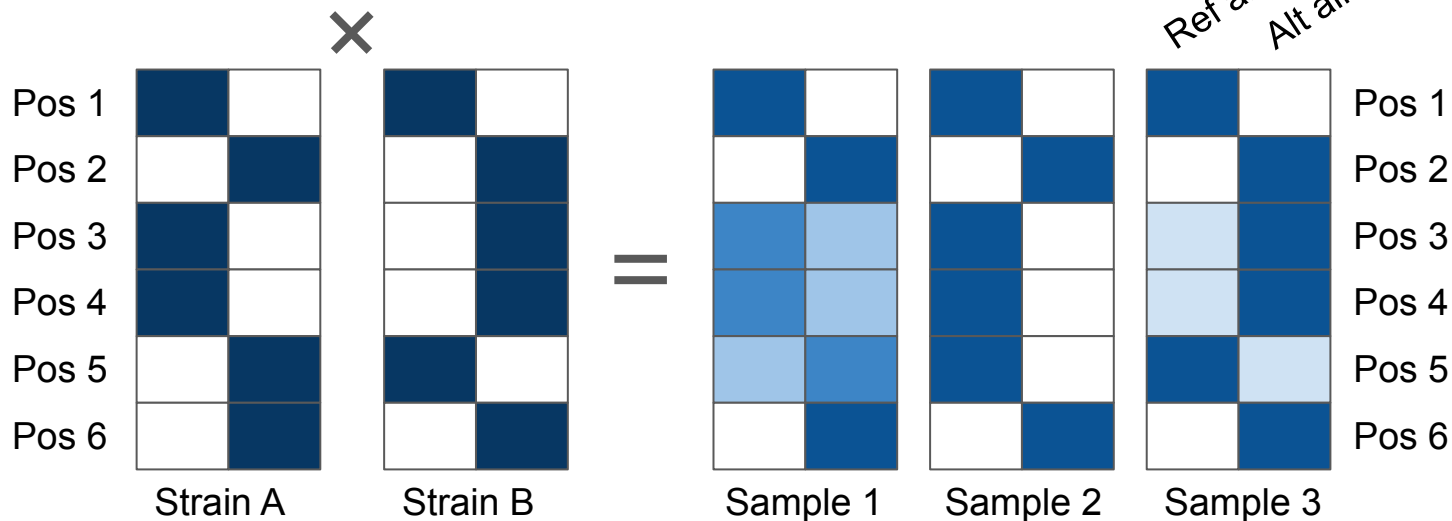
- Most strain diversity has not been previously documented
- Alignment and assembly based methods have been important tools, but require sufficient **sequencing depth** and **strain-pure samples**
- Tools are available for high-throughput
- “metagenotyping”
 - e.g. GT-PRO

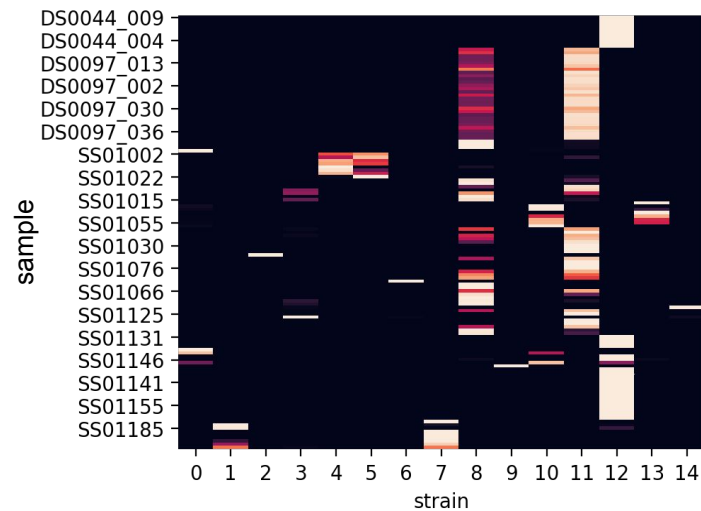
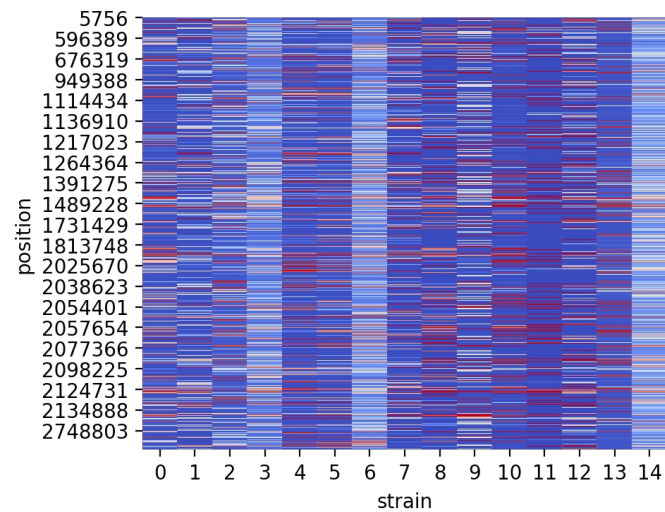
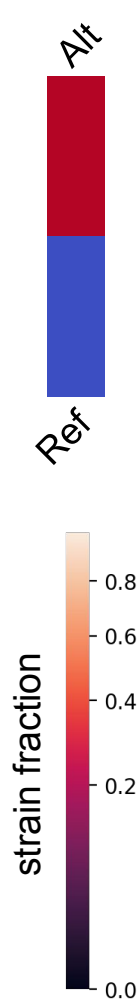
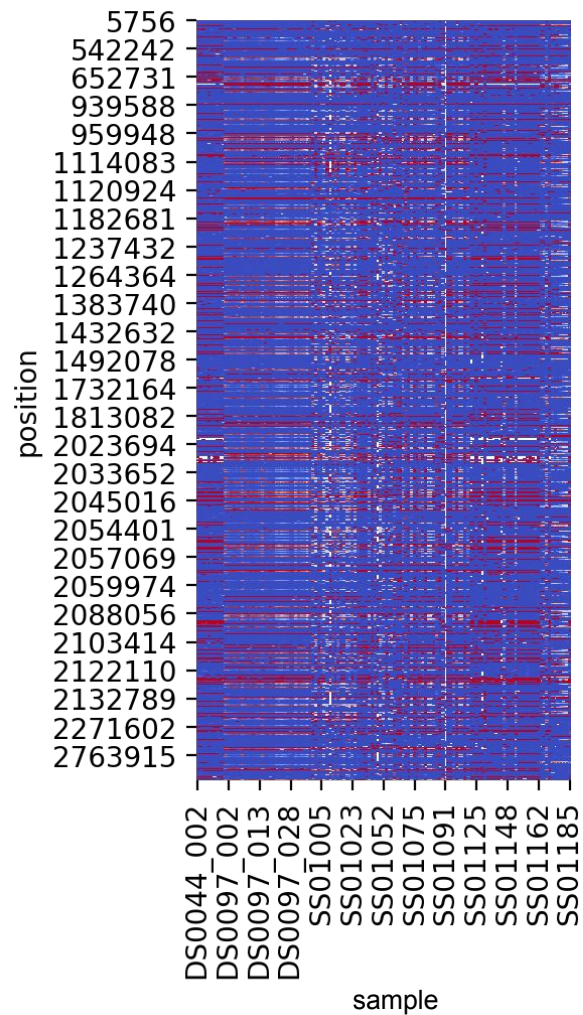
Identifying strains in metagenomes

- Tools are available for high-throughput “metagenotyping”
- **How to recover strain genotype/abundance from metagenotype?**

Species A: Sample 001		
Genome Position	Ref (Count)	Alt (Count)
0001	A (8)	C (1)
0114	T (11)	C (0)
1005	C (0)	G (7)
1202	C (12)	T (5)
...	...	...

Resolving strains from metagenotype matrix





GT-PRO metagenotypes work great

Pros:

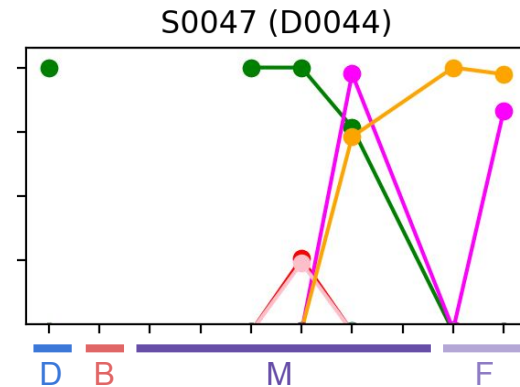
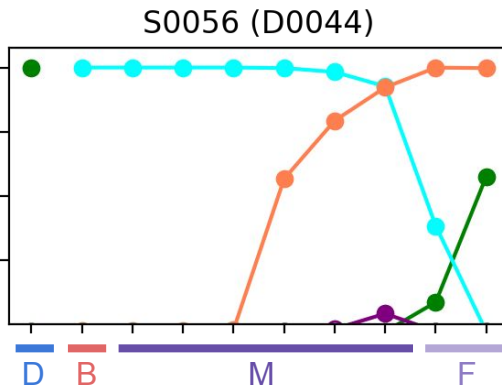
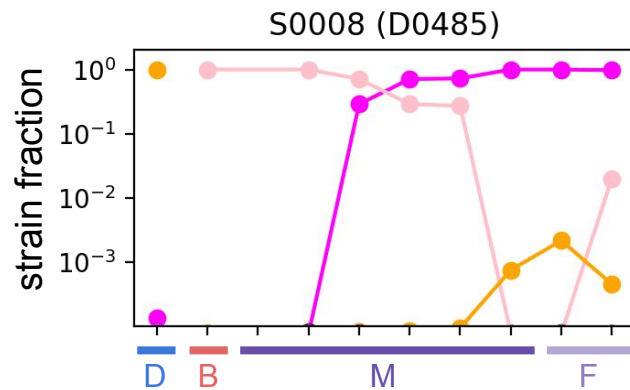
- Preconstructed DB of core genome SNPs
- Fast
- Pre-computed results for >20,000 publicly available metagenomes

Cons:

- Can only use known alleles

We can track strains in an FMT experiment

Strain

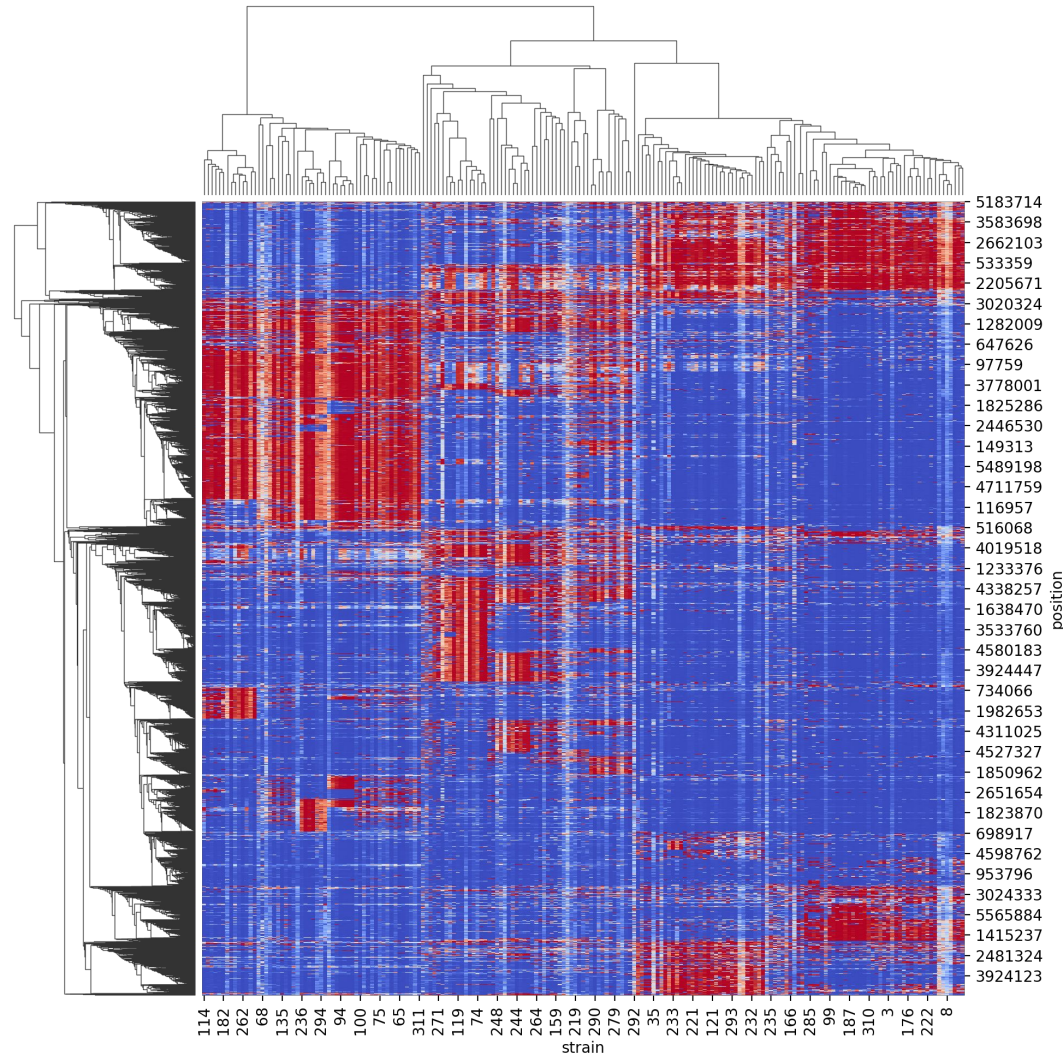


What can we do with this tool?

- Computation (time and memory) scale approximately linearly with dimensions
 - Can be greatly accelerated on GPUs, making it possible to run on thousands of positions, samples, strains
 - GT-PRO metagenotypes were previously collected for 25,133 human metagenomes
-
- *What strains exist?* • *How are they related?* • *How do they evolve?* •

We can identify hundreds of distinct *E. coli* genotypes

- Genotypes cluster into separate sub-types, suggesting incomplete recombination and/or shared evolutionary history



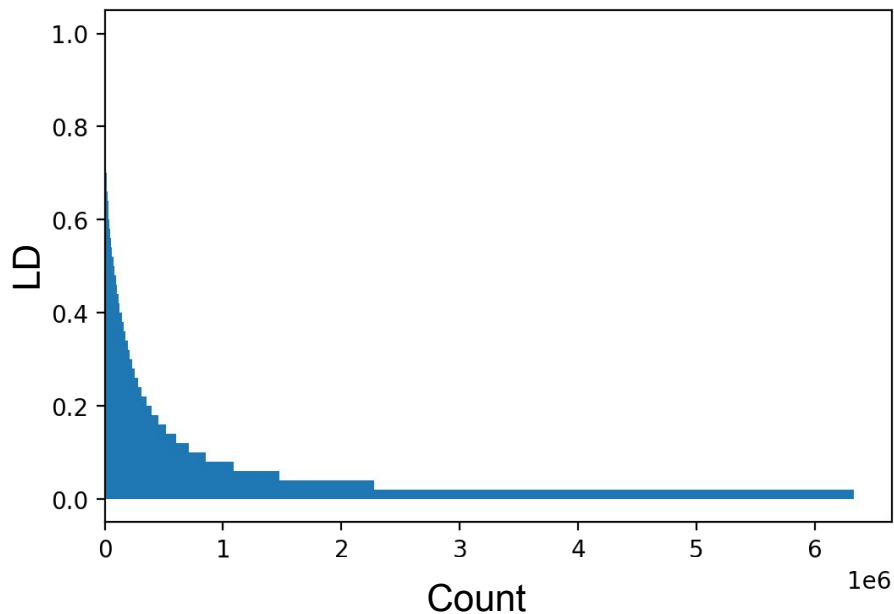
Linkage disequilibrium (LD) among *E. coli* strains

- Two loci are “linked” if the allele at one position is correlated with the allele at the other across strains

		Locus 1	
		A	a
Locus 2	B	0.75	0.25
	b	0.25	0.75

Most alleles are only weakly linked in *E. coli*

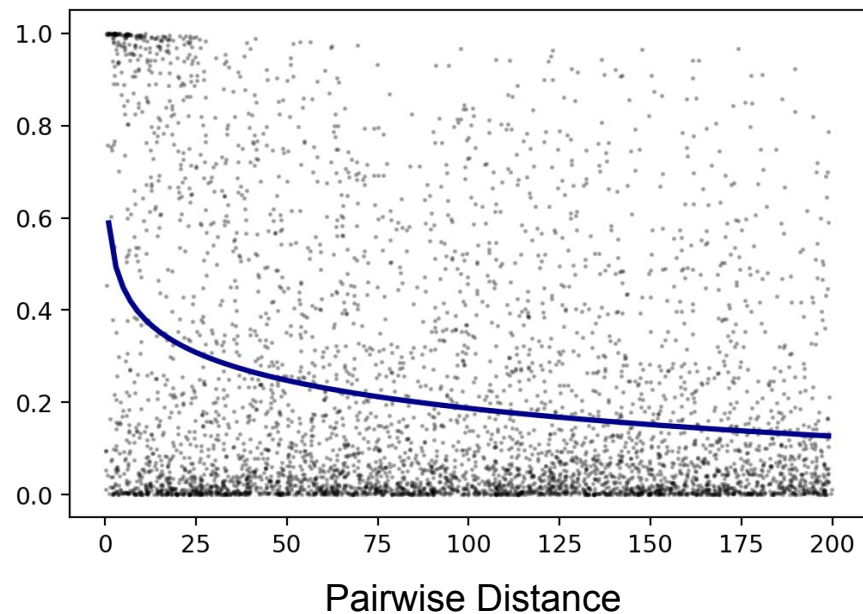
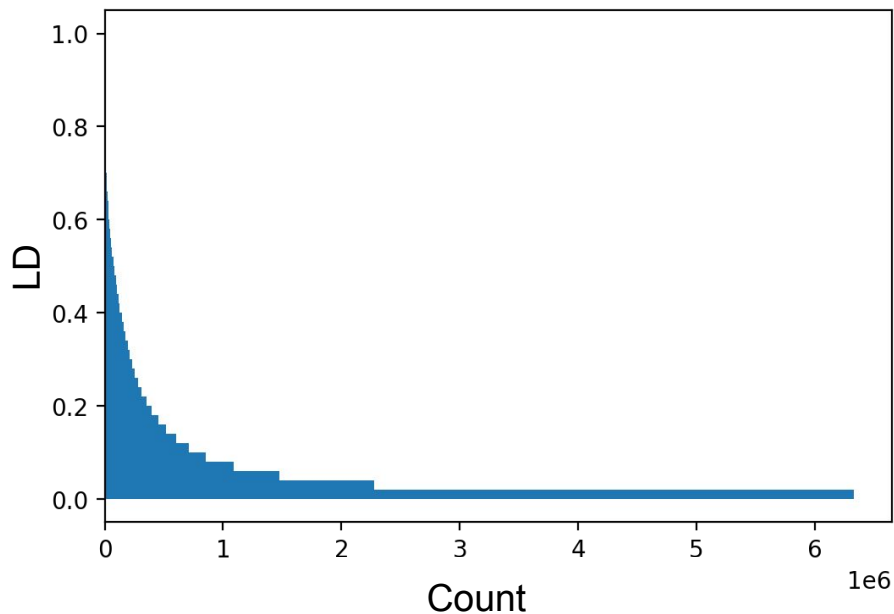
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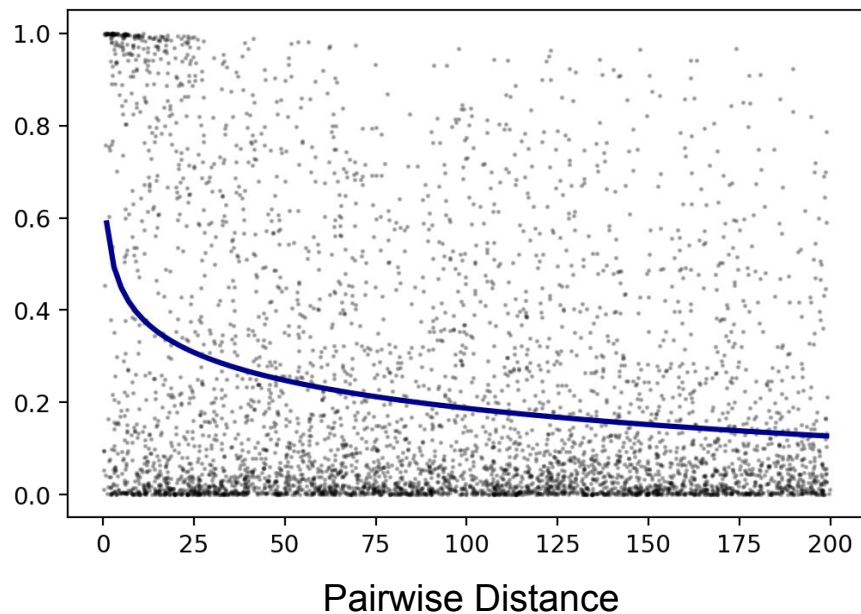
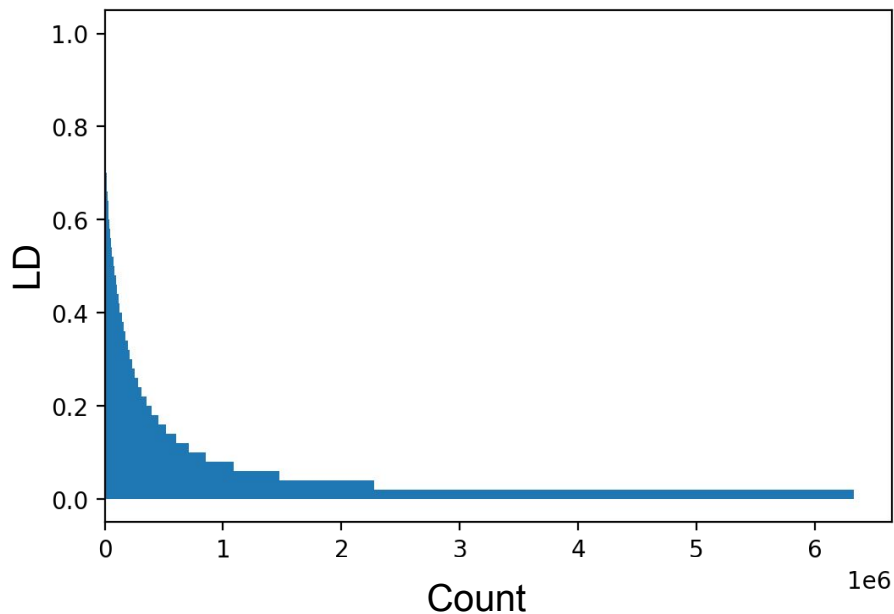
Much higher linkage between nearby alleles

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E. coli LD patterns consistent with high recombination

- Theoretically: with no recombination, linkage is nearly 100% across all pairs
- As recombination increases, linkage decreases overall, but less for proximate pairs



Conclusion

- Strain deconvolution from metagenotypes greatly improves surveys of within-species diversity
- Enables precise strain-tracking in e.g. FMT studies
- Scales well to very large datasets
- Allows us to explore bacterial population structure and evolution